

- 1 M.A.C. Hinton, Br. Mus. Lond. 1 (1926).
- 2 S.I. Ognev, in: Zveri SSSR i prilozhashchikh stran, vol. 7. Acad. Sci. Moscow-Leningrad 1950.
- 3 I.M. Gromov, in: Mlekopitayushchie fauni SSSR, vol. 1. Acad. Sci. Moscow-Leningrad 1963.
- 4 B.S. Yudin, V.G. Krivosheyev and V.G. Belyaev, in: Melkie mlekopitayushchie severa Dalnego Vostoka. Nauka, Novosibirsk 1976.
- 5 M. Seabright, Chromosoma 36, 204 (1972).
- 6 A. Levan, K. Fredga and A.A. Sandberg, Hereditas 52, 201 (1964).
- 7 M.J.D. White, in: Animal cytology and evolution. Cambridge Univ. Press, 1973.
- 8 N.S.F. Ma, T.C. Jones, Folia primat. 24, 282 (1975).
- 9 J.T. Mascarello, J.W. Warner and R.J. Baker, J. Mammal. 55, 831 (1974).
- 10 T. Sharma and R. Raman, Chromosoma 41, 75 (1973).
- 11 A.C. Wilson, G.L. Bush, S.M. Case and M.-C. King, Proc. nat. Acad. Sci. USA 72, 5061 (1975).

'Boost' in gibberellin response by water-stress in seedling growth¹

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Summary. The seedling growth of *Brassica campestris* var. *varuna*, has been studied, as affected by water-stress and gibberellin treatments. A 'boost' in the net GA response due to water-stress, has been observed. Thus presence of GA can overcome the water-stress effects.

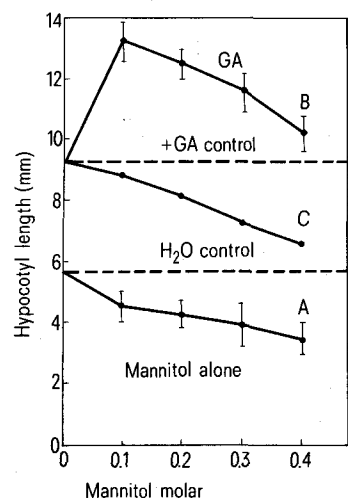
Longitudinal growth caused either by auxin or gibberellin is always associated with increase in water uptake²; the same is true for leaf expansion caused by Kinetin³. Role of this water uptake has been examined in some detail in auxin-induced growth, by use of respiratory inhibitors⁴, and also by providing osmoticum by mannitol, and studying the permeability of the membrane to water⁵, and other solutes⁶. The present experiment was designed to study the interaction of osmoticum provided by mannitol and gibberellic acid (GA)-induced growth in *Brassica campestris* seedlings. It has been noticed that, in presence of mannitol and GA together, both the hormone-induced growth and also the stress-induced inhibition are only partially expressed. So to elaborate their role in such an interaction, the following experiment was performed. Basic methods have been described earlier⁷.

Germinated seeds of *B. campestris* were given 24 h of the stress treatment using the different concentrations of mannitol (0.1–0.4 M) and from each treatment half the seedlings were transferred to water and the other half transferred to GA (10 mg/l) solution, to study the recovery of the stress effects in water or in GA. Growth measurements were taken 48 h after transfer from mannitol solution, and hypocotyl length is the mean of 20 seedlings. A separate control was run for comparison, which comprised of the first 24 h in water and then half the seedlings to water

(water control) and the other half to GA (GA control). Respective mannitol control refers to half the seedlings given stress (0.1–0.4 M) and then transferred to water, as compared to other half transferred to GA.

Hypocotyl length measured after 48 h of recovery is plotted in the figure, where curve A represents the hypocotyl length of seedlings transferred from different mannitol concentrations to water against the water control, thus showing the inhibition caused by the pre-stress treatment. The curve B shows the hypocotyl length of seedlings transferred from different mannitol concentrations to GA, showing a net promotion in GA response, and against GA control, whereas the curve C is plotted as growth of the seedlings transferred from different mannitol concentrations to GA solutions, minus the mannitol alone seedlings, showing that the pre-stress still interferes in GA alone response.

It is clear from the figure that the stress-treatment of the seedlings is reflected in an inhibition even at later periods of the growth (curve A), and also that a stress pre-treatment interferes in the net GA response of a normal seedling (curve C). However, when we plot the growth of the stress-pretreated seedling in GA, a very significant picture arises, since all these points keep values higher than the GA control itself. This shows that the pre-treatment with stress in some way makes the seedlings more responsive to GA, thus a sort of 'boost' is observed in GA response. This boost may also reflect that GA in some way helps in the recovery of the injury caused by mannitol stress. A similar pattern has also been observed in *B. nigra* seedlings.



Showing the hypocotyl growth in length of seedlings of *B. campestris* pretreated with different concentrations of mannitol for 24 h and then transferred to water or GA solution. For explanation of curves see text. Vertical bars show SD. The calculated and table values of significance at 5% levels are 397.589 and 2.10 for treatments, and 1.751 and 2.10 for replications.

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- 2 M.B. Wilkins, Physiology of plant growth and development. McGraw Hill, 1969.
- 3 S. Kuraishi and F.S. Okumura, Bot. Mag. Tokyo 69, 300 (1956); D. Banerji and M.M. Laloraya, Naturwissenschaften 12, 249 (1965).
- 4 D.P. Hacket and K.V. Thimann, Am. J. Bot. 40, 183 (1953).
- 5 R. Cleland and J. Bonner, Plant Physiol. 31, 350 (1956); K.V. Thimann and E.W. Samuel, Proc. nat. Acad. Sci. USA 41, 1029 (1955); L. Ordin and J. Bonner, Plant Physiol. 31, 53 (1956); J.B. Power and E.C. Cocking, J. exp. Bot. 21, 64 (1970).
- 6 L. Reinhold and R.G. Powell, J. exp. Bot. 9, 82 (1958); S. Meylan and P.E. Pillet, Physiol. Veg. 4, 221 (1966); I. Ilan and L. Reinhold, Physiologia Pl. 16, 596 (1963).
- 7 V.K. Rai and M.M. Laloraya, Physiologia Pl. 20, 897 (1967).